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ISOLATION OF NORBERGENIN FROM *SAXIFRAGA STOLONIFERA**

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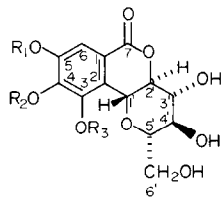
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Key Word Index—*Saxifraga stolonifera*; Saxifragaceae; desmethylbergenin; norbergenin; bergenin; tri-*O*-methyl-norbergenin.

Abstract—Norbergenin, a *C*-glucoside, was isolated from *Saxifraga stolonifera* and its structure assigned as 2 β -*D*-glucopyranosylgallic acid δ -lactone.

Bergenin (**2**), one of the simplest *C*-glucosides, occurs in a number of plants [1–4]. In previous publications [5, 6], its distribution and biosynthesis have been reported. The results of incorporation of [14 C]glucose into **2** by leaves of *Saxifraga stolonifera* showed that gallic acid accepted a glucosyl moiety and was a precursor of **2**. During this study an unidentified phenolic, having high radioactivity, was isolated which was thought to be a demethylation product of **2**; it was named desmethylbergenin (**1**) [6]. While preparing a manuscript on its identification, Kalidhar *et al.* [7] reported the same compound in *Woodfordia fruticosa* (Lythraceae) and called it norbergenin. We now report the confirmation of its structure, as isolated from *S. stolonifera*.



- 1** $R_1 = R_2 = R_3 = H$
Norbergenin
- 2** $R_1 = R_3 = H, R_2 = Me$
Bergenin
- 3** $R_1 = R_2 = R_3 = Me$
Tri-*O*-methylnorbergenin

Our compound and its derivatives were identical in every way with norbergenin and its derivatives as described by Kalidhar *et al.* (see Experimental). In addition, the absolute configuration of **1** was determined by the aromatic chirality method [8, 9]. The three Cotton effects at $\Delta\epsilon_{210} + 2.0$, $\Delta\epsilon_{228} - 12.5$ and $\Delta\epsilon_{290} + 0.8$ showed the chiral lactone ring to be a β -glucosyl moiety of **1**.

Table 1. 13 C NMR chemical shift of norbergenin (**1**), bergenin (**2**) and tri-*O*-methylnorbergenin (**3**)

	1	2	3
Carbon No.	CD ₃ OD-D ₂ O (2:1)	DMSO- <i>d</i> ₆	DMSO- <i>d</i> ₆
1	117.49 <i>s</i>	116.08 <i>s</i>	118.03 <i>s</i>
2	114.08 <i>s</i>	112.76 <i>s</i>	115.93 <i>s</i>
3	143.31 <i>s</i>	142.43 <i>s</i>	148.04 <i>s</i>
4	140.93 <i>s</i>	139.61 <i>s</i>	140.59 <i>s</i>
5	146.58 <i>s</i>	145.94 <i>s</i>	150.92 <i>s</i>
6	111.25 <i>d</i>	109.30 <i>d</i>	109.50 <i>d</i>
7	166.65 <i>s</i>	163.73 <i>s</i>	163.34 <i>s</i>
1'	73.88 <i>d</i>	72.27 <i>d</i>	72.17 <i>d</i>
2'	75.14 <i>d</i>	73.78 <i>d</i>	73.73 <i>d</i>
3'	80.99 <i>d</i>	79.82 <i>d</i>	79.82 <i>d</i>
4'	71.44 <i>d</i>	70.90 <i>d</i>	70.71 <i>d</i>
5'	82.40 <i>d</i>	81.57 <i>d</i>	81.77 <i>d</i>
6'	62.33 <i>t</i>	61.25 <i>t</i>	61.16 <i>t</i>
OMe	{ —	—	61.10 <i>q</i>
	{ —	59.84 <i>q</i>	60.60 <i>q</i>
	{ —	—	56.00 <i>q</i>

*Part 3 in the series "Studies on C-Glycosides in Higher Plants". For Part 2 see Taneyama, M. and Yoshida, S. (1979) *Bot. Mag.* **92**, 69.

21.15 MHz, TMS as int. standard.

EXPERIMENTAL

General methods. Mps were uncorr. TLC was carried out on Si gel and on an Avicel plate (0.25 mm). The following solvents were used for PC and TLC: (A) *n*-BuOH–HOAc–H₂O (4:1:2); (B) 6% HOAc; (C) 70% PhOH; (D) EtOAc–MeOH–H₂O (77:13:10).

Isolation. *Saxifraga stolonifera* was collected at Kanagawa Prefecture, Japan, in Dec. 1978. The whole plants (2.1 kg fr. wt) of *S. stolonifera* were extracted twice with 16 l. hot 80% EtOH. The combined extract was concd under red. pres. to 500 ml and the soln was filtered. The filtrate was extracted with Et₂O and EtOAc successively. The EtOAc-soluble fraction was concd to dryness and then dissolved in 50 ml H₂O. The H₂O-soluble portion was fractionated through the polyamide column (30 × 2 cm, Wako C-200) with 6% HOAc. The fractions containing **1** were concd under red. pres. to a small vol. An aliquot was streaked on paper, which was developed with solvent A followed by solvent B. The band corresponding to **1** was cut out and extracted with 80% EtOH. After concn, the extract was applied to TLC (Avicel plate, solvent A). The band corresponding to **1** was extracted with 80% EtOH. The extract was concd under red. pres. and crystals appeared. Recrystallization from hot H₂O with a small amount of MeOH, gave colourless prisms. Mp 277–278° (mmp 276–277°). Yield 110 mg.

Compound 1. *R_fs* (A, B, C, D) of **1** PC 0.22, 0.40, 0.42, 0.43 (detecting with FeCl₃–K₃Fe(CN)₆ (1:1)). *R_fs* (A, D) on TLC (Si gel) 0.53, 0.93. Purple colour in UV light, blue-purple under UV light + NH₃ vapour. Analysis: calcd for C₁₃H₁₄O₉: C, 49.68; H, 4.49%; found: C, 49.39; H, 4.62%. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 222 (4.27), 289 (3.93); $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOAc}}$ nm: 212, 245 (sh), 338; $\lambda_{\text{max}}^{\text{EtOH} + \text{AlCl}_3}$ nm: 223, 240 (sh), 325; $\lambda_{\text{max}}^{\text{EtOH} + \text{AlCl}_3 + \text{HCl}}$ nm: 222, 289, IR $\nu_{\text{max}}^{\text{Nujol}}$ cm⁻¹: 3470, 3310, 3250 (OH), 2850 (CH₂), 1690 (C=O), 1618, 1595, 1530 (aromatic), 1250 (aromatic C–O) and 860 (C–H). ¹H NMR (100 MHz, DMSO-*d*₆, TMS): δ 3.20–4.04 [6H, C-2'–C-5', C-6' (CH₂)], 4.96 (1H, *d*, *J* = 10.3 Hz, C-1'), 7.13 (1H, *s*, C-6). For ¹³C NMR data see Table 1. CD (H₂O; *c* 0.64). ORD (H₂O; *c* 0.61): $[\phi]_{215} + 15.5 \times 10^5$, $[\phi]_{222} 0$, $[\phi]_{233} - 14.4 \times 10^5$. $[\alpha]_D^{25} - 22.9^\circ$ (H₂O; *c* 0.393).

Demethylation of 2. This was achieved by HI and **1** was obtained. It gave a single spot on PC, *R_fs* (A, B, C, D) 0.22, 0.40, 0.42, 0.43 and TLC (Si gel), *R_fs* (A, D) 0.53, 0.93. Analysis: calcd for C₁₃H₁₄O₉: C, 49.68; H, 4.49%; found: C, 49.20; H, 4.45%. Mp 278°. UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ): 222 (4.35), 289 (3.96). $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOAc}}$ nm: 212, 245 (sh), 338. $\lambda_{\text{max}}^{\text{EtOH} + \text{AlCl}_3}$ nm: 223, 240 (sh), 325. $\lambda_{\text{max}}^{\text{EtOH} + \text{AlCl}_3 + \text{HCl}}$ nm: 222, 289. The results of the IR, ¹H NMR, CD and ORD spectra agreed with those of **1**. ¹³C NMR data are shown in Table 1.

Hydrolysis of 1. The hydrolysis was carried out by the method of ref. [10]. Compound **1** (2 mg) in satd Ba(OH)₂ soln (2 ml) was heated at 100° for 2 hr in a boiling water-bath in a sealed tube.

The gallic acid was isolated from the Et₂O-soluble fraction of the hydrolysate. Gallic acid was confirmed by PC, TLC, UV and IR.

Methylation of 1 and 2. Compounds **1** and **2** were methylated with ethereal CH₂N₂, and tri-*O*-methylnorbergenin was obtained, mp 194–196°. It gave a single spot on PC, *R_fs* (A, B) 0.67, 0.76 and TLC (Si gel), *R_fs* (A, B) 0.80, 0.75. Analysis: Calcd C₁₆H₂₀O₉: C, 53.93, H, 5.66%; found: C, 53.80, H, 5.55%. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 227, 267. $\lambda_{\text{max}}^{\text{MeOH} + \text{NaOAc}}$ nm: 227, 267. $\lambda_{\text{max}}^{\text{MeOH} + \text{AlCl}_3}$ nm: 226, 267. ¹H NMR (100 MHz, DMSO-*d*₆, TMS as int. standard): δ 3.26–4.02 [6H, C-2'–C-5', C-6' (CH₂)], 3.79 (3H, *s*, C-3, OMe), 3.84 (3H, *s*, C-5, OMe), 4.78 (1H, *d*, *J* = 10.7 Hz, C-1'), 7.32 (1H, *s*, C-6). For ¹³C NMR data see Table 1. GC/MS (probe), 70 eV, *m/z* (rel. int.): 572 [M]⁺ (0.07), 557 [M – Me]⁺ (0.3), 500 (50), 499 [M – 73]⁺ (21), 356 (15), 399 (29), 311 (37), 309 (57), 237 (47), 224 (100), 208 (26), 117 (83), 103 (48), 75 (68).

From **2** a dimethyl ether was prepared by the same method which had the same characteristics as **1** trimethyl ether.

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